by Pople and co-workers8 has shown that linear structure 2 is more stable than the bridged one 3. In the course of nitrogen fixation^{9,10}



it is highly unlikely that proton transfer to free nitrogen occurs. Instead, such proton transfer probably occurs on to coordinated dinitrogen on a suitable transition-metal site such as molybdenum. In continuation of our studies on substituted diazonium ions (such as $N_2^+NH_2$, 11 $N_2^+NO_2$, 12 N_2^+CN , 12 $N_2^+F^{12}$, etc.) we would like to report new results on our attempts to generate diazonium ion 1 not by direct protonation but by diazotization of ammonia and some of its derivatives using nitrosonium tetrafluoroborate salt.

It is known that aromatic as well as aliphatic amines and isocyanates react with NO+ salts, such as NO+BF₄- to form the corresponding diazonium ions.¹³ Weiss recently reported¹⁴ a useful

$$RNH_2 + NO^+BF_4^- \rightarrow RN^+ \equiv NBF_4^- + H_2O$$

 $RN=C=O + NO^+BF_4^- \rightarrow RN^+ \equiv NBF_4^- + CO_2$

additional anhydrous diazotization method by reacting bissilylated amines with NO+ salt.

$$R - N(Si(CH_3)_3)_2 + NO^+X^- \rightarrow RN^+ \equiv N X^- + [(CH_3)_3Si]_2O$$

As the direct protonation of nitrogen could not be achieved in solution, it occurred to us that the problem could be attacked by generating protonated dinitrogen, i.e., the parent diazonium ion via diazotizing ammonia and some of its derivatives.

Using 96% enriched ¹⁵NO⁺BF₄⁻ salt¹⁵ we have indeed succeeded with diazotization of ammonia, bis(trimethylsilyl)amine, and isocyanic acid, respectively, resulting in the formation of ¹⁴N=¹⁵N. The mono ¹⁵N-labeled nitrogen gas obtained can be produced only through the intermediacy of the parent diazonium ion 1.

All reactions were carried out in dichloromethane solution and the nitrogen evolved during the reaction was analyzed by GC-

(9) Chatt, J.; Dilworth, J. R.; Richards, R. L. Chem. Rev. 1978, 78, 589. Also see: Bottomley, F., Burns, R. C., Eds. "Treatise on Dinitrogen Fixation";

Chem. Soc. 1983, 105, 5657.

(12) Olah, G. A.; Laali, K.; Farnia, M.; Shih, J.; Singh, B. P.; Christe, K.
O.; Schack, C. J. J. Org. Chem. 1985, 50, 1338.
(13) (a) Kirmse, W. Angew. Chem., Int. Ed. Engl. 1976, 15, 251. (b)
Zollinger, H. "Azo and Diazo Chemistry"; Wiley-Interscience: New York,

(14) Weiss, R.; Wagner, K.-G.; Hertel, M. Chem. Ber. 1984, 117, 1965. (15) 96% ¹⁵N enriched NO⁺BF₄ was prepared by treating 96% ¹⁵NOCl with HF-BF₃ in nitromethane solution at -78 °C. The 96% ¹⁶NOCl was generated by treating 96% Na15NO2 with dry HCl in nitromethane solvent with CaCl2 drying.

MS. 16 Ammonia itself reacts with nitrosonium tetrafluoroborate rather violently even at -80 °C to give 89% monolabeled and 11% unlabeled dinitrogen. In reaction of bis(trimethylsilyl)amine 94% monolabeled nitrogen was formed, the reaction being somewhat sluggish at -80 °C. Isocyanic acid reacted very slowly with the nitrosonium ion even at room temperature. A competing polymerization of isocyanic acid seem to occur along with the diazotization and it was not possible to determine the exact isotope distribution of the evolved nitrogen although it is estimated to be ≥90% monolabeled.

Attempts were made to directly detect the monolabeled diazonium ion 1 using ¹⁵N NMR spectroscopy. ¹⁷ In a typical experiments ≈50 mg of bis(trimethylsilyl)amine was treated with ≈ 100 mg of 95% ¹⁵NOBF₄ in 2 mL of dichloromethane in a 10-mm NMR tube at -80 °C in the NMR probe. The probe temperature when raised to -40 °C resulted in slow evolution of nitrogen gas. Accumulation of ¹⁵N data over a period of 30 min detected, however, the presence of only ¹⁵NO⁺ salt and no signal for 1 could be observed. The failure to detect HN2+ in the described stop-flow experiment seems to indicate that 1 is unstable under the reaction conditions. This is in accordance with the known low proton affinity of dinitrogen.

Our studies reported indicate that the parent diazonium ion 1 was in situ formed in the diazotization of ammonia and its derivatives with ¹⁵NO⁺BF₄. However, 1 once formed has very short lifetime to be observed by NMR spectroscopy and spontaneously deprotonates to $^{15}N \stackrel{14}{\equiv} ^{14}N$.

Acknowledgment. Support of our work by the National Science Foundation is gratefully acknowledged. R. Herges is grateful for the financial support by a Feodor Lynen grant of the Alexander von Humbold Foundation.

(16) GC-MS analysis was performed on a Hewlett-Packard 5985 A GC-

MS spectrometer.
(17) The ¹⁵N NMR studies were carried out on a Varian Associates Model FT-80 NMR spectrometer equipped with a variable-temperature broad-band

(18) Bohme, D. K.; Makay, G. I.; Shiff, U. H. I. J. Chem. Phys. 1980, 73,

Total Synthesis of the Cytochrome P-450 Epoxygenase Metabolites 5(R),6(S)-,5(S),6(R)-, and 14(R),15(S)-Epoxyeicosatrienoic Acid (EET) and Hydration Products 5(R), 6(R) and 14(R), 15(R)-Dihydroxyeicosatrienoic Acid (DHET)¹

Christine A. Moustakis, Jacques Viala, Jorge Capdevila, and J. R. Falck*

> Departments of Molecular Genetics and Biochemistry University of Texas Health Science Center at Dallas Dallas, Texas 75235 Received May 9, 1985

Recent reports³ have elucidated an alternative mode of eicosanoid production,4 designated the epoxygenase pathway,5a that

(3) Reviews: Capdevila, J.; Saeki, Y.; Falck, J. R. Xenobiotica 1984, 14, 105-118. Yamamoto, S. In "New Comprehensive Biochemistry"; Pace-Asciak, C., Granstrom, E., Eds.; Elsevier: New York, 1983; Vol. 5, Chapter 5, pp 171-202.

(4) For metabolism of other fatty acids, see: Van Rollins, M.; Baker, R. C.; Sprecher, H. W.; Murphy, R. C. J. Biol. Chem. 1984, 259, 5776-5783. Oliw, E. H. Biochem. Biophys. Res. Commun. 1983, 111, 644-651. A related pathway in plants has been described: Kato, T.; Yamaguchi, Y.; Hirano, T.; Yokoyama, T.; Uyehara, T.; Namai, T.; Yamanaka, S.; Harada, N. Chem. Lett. 1984, 409-412 and references cited therein.

⁽⁸⁾ Whitesides, R. A.; Frisch, M. J.; Binkley, J. S.; DeFrees, D. J.; Schlegel, H. B.; Ragavachari, K.; Pople, J. A. "Carnegie-Mellon Quantum Chemistry Archive", 2nd ed.; Carnegie-Mellon University: Pittsburgh, 1981. For related calculations on frequencies and IR intensities of 1, see: Botschwina, P. Chem. Phys. Lett. 1984, 107, 535.

⁽¹⁾ Presented in part: Moustakis, C. A.; Viala, J.; Falck, J. R. "Abstracts of Papers", 189th National Meeting of the American Chemical Society, Miami Beach, FL, April-May, 1985; American Chemical Society: Washington, DC, 1985; ORGN-174.

(2) NATO postdoctoral fellow.

^a NaIO₄, MeOH/H₂O 2:1, -15 → 0 °C, 1.5 h. ^b BuPPh₃, BuLi, THF/HMPA 4:1, -78 → 0 °C, 12 h. ^c 5% Pd/C, 1 atm H₂, EtOAc/MeOH 1:1, 1 h. ^d KH, ^tBuPh₂SiCl, THF, 12 h. ^e AcOH/THF/H₂O 5:2:2, 60-65 °C, 2-2.5 h. ^f 8, LiN (SiMe₃)₂, THF/HMPA, -78 → 23 °C, 12 h. ^g Bu₄NF, THF, 0 °C, 24 h. ^h TsCl, py, DMAP, CH₂Cl₂, 23 °C, 48 h.

is distinct from the cyclooxygenase and lipoxygenase branches of the arachidonate cascade. Initial metabolism is mediated by cytochrome P-450 and leads to four novel, regioisomeric cis-epoxyeicosatrienoic acids⁶ (EETs) as well as lipoxygenase-type Z,E dienols⁷ and $\omega/(\omega-1)$ oxygenated products.⁸ The EETs exhibit significant in vitro biological activities, inter alia, stimulation of peptide hormone release,⁵ mobilization of microsomal calcium,⁹ and alteration of net potassium and sodium flux in the isolated rabbit kidney tubule.¹⁰ Hydration of the EETs by epoxide hydrolases¹¹ results in vic-dihydroxyeicosatrienoic acids (DHETs) which also may have a physiological role.^{5a,b} These observations,

in conjunction with the in vivo detection of EETs in mammalian tissue, ¹² implicate the cytochrome P-450 epoxygenase pathway as a potential participant in homeostasis. Due to the minute amounts of epoxygenase metabolites isolable from natural sources, we report herein the convergent, enantiospecific total synthesis of the title compounds utilizing an easily obtainable carbohydrate precursor.

The strategy summarized in Scheme I was calculated to provide access to both DHETs and EETs from a single chiral intermediate. Methyl furanoside (1), available¹³ as an \sim 4:1 anomeric mixture from 2-deoxy-D-glucose (0.1% HCl, MeOH, 2 h; neutralization with excess BaCO₃), was converted to the pivotal aldehyde 2¹⁴ by NaIO₄ cleavage in 71-73% yield after chromatography. Elaboration using butyltriphenylphosphorane and catalytic reduction of the resultant olefin furnished 3 (92%) which was transformed to lactol 4 (71%) by silylation (t-BuPh, SiCl, KH, THF, 12h) and hydrolysis in AcOH/THF/ H_2O (5:2:2, 60-65 °C, 2-2.5 h). Condensation of 4 with the ylide (3.3 equiv) derived from [10carbomethoxydeca-(Z,Z)-3,6-dien-1-yl]triphenylphosphonium bromide¹⁵ (8) in 4:1 THF/HMPA (-78 to 23 °C over 12 h) afforded 5 and a small amount of the corresponding 11-E isomer in 50-53% yield. Exposure of 5 to Bu₄NF smoothly generated (95%) methyl 14(R), 15(R)-dihydroxyeicosatrienoate¹⁶ (6).

Tosylation of **5**, fluoride anion desilylation with concomitant epoxide closure, gave methyl 14(R), 15(S)-epoxyeicosatrienoate¹⁶ (7) (64% from **5**), the predominant antipode of 14,15-EET produced by incubation of arachidonic acid with the major phenobarbital-inducible form of rat liver microsomal cytochrome P- $450.^{17}$ The stereochemical homogeneity of 7 was verified following olefin reduction (5% Pd/C, EtOAc, 1 atm H₂) by NMR analysis using chiral lanthanide shift reagents¹⁸ under conditions which fully resolve a d,l mixture.

In contrast to its otherwise high enantiofacial selectivity, purified microsomal cytochrome P-450 produces 5,6-EET as a 60:40 enantiomeric mixture.¹⁷ It was of interest, therefore, to prepare both isomers for independent pharmacological evaluation (Scheme II). Three-carbon homologation of 2 with [3-bis(isopropoxy)-propyl]triphenylphosphonium bromide¹⁹ (16) and hydrogenation over 5% Pd/C furnished 9 (65%). Selective hydrolysis (0.4 M HCO₂H) of the isopropyl acetal in the presence of *m*-chloroperbenzoic acid gave the corresponding carboxylic acid. In situ esterification (Me₂SO₄, NaHCO₃, 23 °C, 10 h) and extractive isolation provided methyl ester 10 from which lactol 11 was obtained by acidic hydrolysis (0.5 M HCO₂H, 65 °C) in 79% yield from 9. Union of 11 under Wittig cis-olefination conditions

^{(5) (}a) Capdevila, J.; Chacos, N.; Falck, J. R.; Manna, S.; Negro-Vilar, A.; Ojeda, S. R. Endocrinology 1983, 113, 421-423. (b) Snyder, G. D.; Capdevila, J.; Chacos, N.; Manna, S.; Falck, J. R. Proc. Natl. Acad. Sci. U.S.A. 1983, 80, 3504-3507. (c) Falck, J. R.; Manna, S.; Moltz, J.; Chacos, N.; Capdevila, J. Biochem. Biophys. Res. Commun. 1983, 114, 743-749. (d) Negro-Vilar, A.; Snyder, G. D.; Falck, J. R.; Manna, S.; Chacos, N.; Capdevila, J. Endocrinology 1985, 116, 2663-2668.

⁽⁶⁾ Falck, J. R.; Manna, S. Tetrahedron Lett. 1982, 23, 1755-1756. Chacos, N.; Falck, J. R.; Wixtrom, C.; Capdevila, J. Biochem. Biophys. Res. Commun. 1982, 104, 916-922. Oliw, E. H.; Guengerich, F. P.; Oates, J. A. J. Biol. Chem. 1982, 257, 3771-3781.

⁽⁷⁾ Falck, J. R.; Siddhanta, A. K.; Estabrook, R. W.; Capdevila, J.; Mioskowski, C. Tetrahedron Lett. 1984, 25, 1457-1458. Capdevila, J.; Marnett, L. J.; Chacos, N.; Prough, R. A.; Estabrook, R. W. Proc. Natl. Acad. Sci. U.S.A. 1982, 79, 767-770.

⁽⁸⁾ Manna, S.; Falck, J. R.; Chacos, N.; Capdevila, J. Tetrahedron Lett. 1983, 24, 33-36 and references cited therein.

⁽⁹⁾ Kutsky, P.; Falck, J. R.; Weiss, G. B.; Manna, S.; Chacos, N.; Capdevila, J. *Prostaglandins* 1983, 26, 13-21.

⁽¹⁰⁾ Jacobson, H. R.; Corona, S.; Capdevila, J.; Chacos, N.; Manna, S.; Womack, A.; Falck, J. R. In "Prostaglandins and Membrane Ion Transport"; Braquet, P., Frolich, J. C., Nicosia, S., Garay, R., Eds.; Raven Press: New York, 1984; pp 311-318.

⁽¹¹⁾ Chacos, N.; Capdevila, J.; Falck, J. R.; Manna, S.; Martin-Wixtrom, C.; Gill, S. S.; Hammock, B. D.; Estabrook, R. W. Arch. Biochem. Biophys. 1983, 223, 639-648.

⁽¹²⁾ Capdevila, J.; Pramanik, B.; Napoli, J. L.; Manna, S.; Falck, J. R. Arch. Biochem. Biophys. 1984, 231, 511-517.

⁽¹³⁾ Hughes, I. W.; Overend, W. G.; Stacey, M. J. Chem. Soc. 1949, 2846-2849.

⁽¹⁴⁾ All new compounds had satisfactory carbon/hydrogen microanalyses (±0.3%) and/or high-resolution mass spectra.

⁽¹⁵⁾ Prepared in 64% overall yield from 4-[(tert-butyldiphenylsilyl)-oxy]-1-bromo-2-butyne by CuI (1.7 equiv) catalyzed coupling with the dianion of 5-hexynoic acid (1.7 equiv, EtMgBr) in THF/HMPA (5:1) followed by in situ esterification with Me₂SO₄/NaHCO₃. Sequential reduction of the resultant diyne over P-2 nickel [NaBH₄, Ni(OAO₂), H₂NCH₂CH₂NH₃, EtOH, 1 atm of H₂], desilylation (20% ω/ω camphorsulfonic acid, MeOH), alcohol/bromide interchange (Ph₃P, CBr₄, CH₂Cl₂), and displacement with triphenylphosphine (CH₃CN, 85 °C) gave 8. For an alternative sequence, see: Nicolaou, K. C.; Hernandez, P. E.; Ladduwahetty, T.; Randall, J. L.; Webber, S. E.; Li, W. S.; Petasis, N. A. J. Org. Chem. 1983, 48, 5404–5406.

⁽¹⁶⁾ The corresponding free acid was obtained quantitatively by basic hydrolysis (LiOH, MeOH/H₂O 3:1, 3 h), eareful acidification to pH 4.5, and extractive isolation. Ester and free acid were spectrally and chromatographically (HPLC, TLC) indistinguishable from authentic racemic standards prepared according to: Corey, E. J.; Niwa, H.; Falck, J. R. J. Am. Chem. Soc. 1979, 101, 1586–1587. Reference 8.

⁽¹⁷⁾ Falck, J. R.; Manña, S.; Jacobson, H. R.; Estabrook, R. W.; Chacos, N.; Capdevila, J. J. Am. Chem. Soc. 1984, 106, 3334-3336.

⁽¹⁸⁾ Cockerill, A. F.; Davies, G. L. O.; Harden, R. C.; Rackham, D. M. Chem. Rev. 1973, 73, 553-588. Hart, H.; Love, G. M. Tetrahedron Lett. 1971, 625-628. Manni, P. E.; Howie, G. A.; Katz, B.; Cassady, J. M. J. Org. Chem. 1972, 37, 2769-2771.

⁽¹⁹⁾ Battersby, A. R.; Buckley, D. G.; Staunton, J.; Williams, P. J. J. Chem. Soc., Perkin Trans. 1 1979, 2550-2558.

Scheme IIa

 a 16, BuLi, THF/HMPA 4:1, -78 \rightarrow 23 °C, 12 h. b 5% Pd/C, 1 atm H₂, EtOH, 2 h. 'HCO₂H, MCPBA, THF/H₂O 3:1, 35 h; Me₂S, 30 min. ^d Me₂SO₄, NaHCO₃, 23 °C, 10 h. 'HCO₂H, THF/H₂O 1:1, 65 °C, 2.5 h. ^f 17, LiN(SiMe₃)₂, THF/HMPA, $4:1, -7\$ \rightarrow 23$ °C, 12 h; MeOH, 2 h. g TsOH, PhCH₃, 3 Å Mol. Sieves, reflux, 0.5 h. h TsCl, py, DMAP, CH₂Cl₂, 12-48 h. Et₃N, MeOH, 12 h. j DHP, PPTS, CH₂Cl₂, 12 h. k Amberlyst H-15, MeOH, 12 h. l NaOMe, MeOH, 0 °C, 1 h.

(THF/HMPA 4:1, -78 to 23 °C over 12 h) with the ylide from dodeca-(Z,Z)-3,6-dien-1-yltriphenylphosphonium bromide²⁰ (17), anhydrous methanol quench (23 °C, 2 h), and chromatography secured methyl 5(R), 6(R)-dihydroxyeicosatrienoate¹⁶ (12) (37%). Differentiation of the diol by lactonization to 13 (98%), tosylation, and treatment with $Et_3N/MeOH$ afforded methyl 5(R), 6(S)epoxyeicosatrienoate (14)16 (55%).

Lactone 13 was also exploited for the preparation of methyl 5(S), 6(R)-epoxyeicosatrienoate¹⁶ (15) by the sequence: tetrahydropyranylation, lactone methanolysis, tosylation, THP removal, and epoxide closure under the influence of NaOMe (42% from 13)

The foregoing syntheses provide ready access to sufficient quantities of the epoxygenase metabolites for pharmacological and biological testing. Investigations into their possible physiological role and metabolic fate will be reported in due course.

Acknowledgment. This work was supported financially by the USPHS NIH (GM 31278, AM 34056), a grant-in-aid from the American Heart Association with funds contributed in part by the Texas Affiliate, and NATO (85/026). We thank Dr. Yuh-Lin Yang for experimental assistance and Drs. Charles Mioskowski and A. C. Oehlschlager for their generous advice.

Supplementary Material Available: Chromatographic, microanalytical, and spectral data for 2-7, 9, 10, 12, 14, and 15 (2 pages). Ordering information is given on any current masthead page.

Stabilization of the Phenyl Cation by Hyperconjugation[†]

Yitzhak Apeloig* and Dorit Arad

Department of Chemistry Technion-Israel Institute of Technology Haifa 32000, Israel Received April 1, 1985

Aryl cations 1, have attracted considerable experimental^{1,2} and theoretical interest.^{3,4} In solution, 1 can be generated by the decomposition of arenediazonium ions, 1 but the numerous attempts to generate these species by the solvolysis of aryl precursors have failed.2 These failures result from the inherent low stability of the phenyl cation (1a), which in the gas phase is 21-25 kcal/mol less stable than the 2-propenyl cation (2).⁵ Ab initio calculations^{6,7}

give an energy difference of 27 kcal/mol at MP2/6-31G*.8a,b 2 is among the least stable vinyl cations that can be generated by

[†] Dedicated to Professor David Ginsburg on the occasion of his 65th birthday

(1) (a) Zollinger, H. In "The Chemistry of Triple-bonded Functional Groups"; Patai, S., Rappoport, Z., Eds.; Wiley: London, 1983; Suppl. C. (b) Szele, I.; Zollinger, H. Helv. Chim. Acta 1981, 64, 2728 and references therein. (c) Fornarini, S.; Speranza, M. J. Chem. Soc., Perkin Trans. 2 1984, 171. (d) Speranza, M.; Keheyan, Y.; Angelini, G. J. Am. Chem. Soc. 1983, 105, 6377. (e) Depke, G., Hanack, M., Hummer, W., Schwarz, H. Angew Chem., Int. Ed. Engl. 1983, 22, 786. (f) Haberfield, P.; Cardona, C.; Bell, M. J. Org. Chem. 1984, 49, 78

(2) (a) Streitwieser, A.; Dafforn, A. Tetrahedron Lett. 1976, 1435. (b) Subramanian, L. R.; Hanack, M.; Chang, L. W. K.; Imhoff, M. A.; Schleyer, P. v. R.; Effenberger, F.; Kurtz, W.; Stang, P. J.; Dueber, T. E. J. Org. Chem. 1976, 41, 4099. (c) Laali, K.; Szele, I.; Yoshida, K. Helv. Chim. Acta 1983, 66, 1710. (d) Note, however, the successful generation of a phenyl cation via the solvolytic cyclization of a dienynyl triflate: Hanack, M.; Holweger, W. J. Chem. Soc., Chem. Commun. 1981, 713.

(3) (a) Dill, J. D.; Schleyer, P. v. R.; Binkley, J. S.; Seeger, R.; Pople, J. A.; Haselbach, E. J. Am. Chem. Soc. 1976, 98, 5428. (b) Dewar, M. J. S.; Reynolds, C. H. J. Am. Chem. Soc. 1982, 104, 3244. (c) Schleyer, P. v. R.; Kos, A. J.; Raghavaohari, K. J. Chem. Soc., Chem. Commun. 1983, 1296.

(4) Gleiter, R.; Hoffmann, R.; Stohrer, W. D. Chem. Ber. 1972, 105, 8. (5) (a) Relative to the corresponding hydrocarbons. (b) Based on $\Delta H_i^{\circ}(C_6H_5^+)=270$ kcal/mol: Beauchamp, J. L. Adv. Mass. Spectrom. 1974, 6, 717. A lower value of $\Delta H_i^{\circ}(C_6H_5^+)=266$ kcal/mol was also reported: Sergeev, Y. L.; Akopyan, M. E.; Vilesov, F. I.; Kleimenov, V. I. Opt. Spectrosc. (Engl. Transl.) 1970, 29, 63. $\Delta H_f^{\circ}(2) = 230 \text{ kcal/mol}$: Aue, D. H.; Bowers, M. T. In "Gas Phase Ion Chemistry"; Bowers, M. T., Ed.; Academic Press: New York, 1979; Vol. 2, Chapter 9. $\Delta H_1^{\circ}(C_h H_h) = 19.8$ kcal/mol, $\Delta H_1^{\circ}(CH_2 = CHCH_3) = 4.9$ kcal/mol. Cox, J. D.; Pilcher, G. "Thermochemistry of Organic and Organometallic Compounds"; Academic Press: New York, 1970.

J. A.; Pietro, W. J.; Hehre, W. J. Ibid. 1982, 104, 2797.

(8) (a) 6-3 | G*: Hariharan, P. C.; Pople, J. A. Theor. Chim. Acta 1973, 28, 213. Pietro, W. J.; Francl, M. M.; Hehre, W. J.; DeFrees, D. J.; Pople, J. A.; Binkley, J. S. J. Am. Chem. Soc. 1982, 104, 5039. (b) MP2: Frisch, M. J., Krishnan, R., Pople, J. A. Chem. Phys. Lett. 1980, 75, 66 and references therein. (c) Due to the size of 3 we could not carry out a 6-31G* calculation. We have therefore constructed a smaller fully polarized 3-21G* basis set by adding a set of six d-type functions taken from 6-31G*. We follow the guidelines of in ref 8a. In cases where 6-31G* and 3-21G* calculations are both available the results were uniformly close. A similarly constructed 3-21G* basis set was recently used by: Bachrach, S. M.; Streitwieser, A. Ibid. A similarly constructed 1985, 107, 1186.

⁽²⁰⁾ Obtained in 60-65% overall yield from 2-octyn-1-ol by modification of the procedures in ref 15.